

Engineering Oocytes from Somatic Cells

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Abstract

In vitro gametogenesis (IVG)—the generation of functional gametes from pluripotent or somatic cells in culture—represents a paradigm-shifting frontier in reproductive medicine with the potential to address absolute infertility (Saitou & Hayashi, 2021). This review synthesizes the current state and formidable challenges of human oocyte IVG. We detail the two primary strategies: the stepwise differentiation of induced pluripotent stem cells (iPSCs) through a primordial germ cell-like cell (PGCLC) intermediate, and the direct reprogramming of somatic cells. Both pathways confront significant biological barriers, including the achievement of complete epigenetic reprogramming and correct genomic imprinting, the faithful completion of meiosis I to prevent aneuploidy, and the acquisition of cytoplasmic competence (Seisenberger et al., 2012; Nagaoka, Hassold, & Hunt, 2012). Critically, oocyte development is inseparable from its somatic niche; thus, advances in ovarian organoids and 3D biofabrication are essential to reconstruct the follicular microenvironment (Laronda et al., 2017; Sun, Parikh, & Fuller, 2022). Beyond the laboratory, IVG raises profound ethical and social questions concerning reproductive longevity, embryo selection, the redefinition of parenthood, and equitable access (Greely, 2016; Mathews, Morain, & Finkelstein, 2017). We conclude that while mouse models prove principle, translation to humans requires solving fundamental biological puzzles and concurrently developing rigorous ethical and regulatory frameworks. The clinical application of IVG, likely decades away, must be guided by a paramount commitment to offspring safety and inclusive public discourse.

Keywords: In Vitro Gametogenesis; Human Oocyte; Induced Pluripotent Stem Cells; Epigenetic Reprogramming; Ovarian Organoid; Reproductive Ethics; Genomic Imprinting; Infertility; Bioengineering.

Introduction

Human fertility is constrained by a finite, non-renewable endowment of germ cells. Conditions like premature ovarian insufficiency, gonadotoxic therapies, genetic disorders, and age-related depletion result in an inability to produce functional gametes—an unresolved challenge in reproductive medicine (Choi, Chung, & Lee, 2020). While assisted reproductive technologies (ART) like in vitro fertilization (IVF) offer solutions, they are limited by existing ovarian reserve or introduce complex genetic and ethical issues via donor programmes (Cobo, García-Velasco, & Domingo, 2021). This fundamental gap has driven a paradigm shift towards in vitro gametogenesis (IVG): the differentiation of pluripotent stem cells (PSCs) or somatic cells into functional haploid gametes entirely ex vivo (Saitou & Hayashi, 2021).

Landmark murine studies have demonstrated proof-of-concept, with IVG-derived oocytes yielding live, fertile offspring (Hikabe et al., 2016). However, translating these protocols to human cells has proven vastly more complex, exposing critical knowledge gaps in human germ cell development. This article synthesizes recent breakthroughs in generating human oocytes in vitro, critically analyses persistent technological hurdles—focusing on epigenetic fidelity, meiotic completion, and niche recapitulation—and confronts the unprecedented ethical, social, and regulatory challenges precipitated by this transformative technology.

The Architectures of Creation: Strategic Pathways to an Artificial Oocyte

Two primary, converging strategic pathways are pursued for human IVG: differentiation from pluripotent stem cells and direct reprogramming of somatic cells (Figure 1).

Figure 1: Strategic Pathways for In Vitro Oogenesis

The schematic compares the two main approaches: the Pluripotent Stem Cell (PSC) Route (stepwise differentiation recapitulating development) and the Direct Reprogramming Route (transdifferentiation bypassing pluripotency).

The Pluripotent Stem Cell Route: Recapitulating Development

This established approach guides human ESCs or iPSCs through developmental stages mirroring fetal germ cell development. The first critical step is the induction of primordial germ cell-like cells (PGCLCs) by activating key signaling pathways (e.g., BMP4) and transcription factors like PRDM1, TFAP2C, and SOX17 (Irie et al., 2015; Sasaki et al., 2015). The resulting PGCLCs initiate epigenetic reprogramming, including global DNA demethylation (Seisenberger et al., 2012; Guo et al., 2015). Subsequent progression to oogonia and entry into meiosis requires pro-meiotic signals like retinoic acid and factors such as DAZL (Chen et al., 2014). A major bottleneck is achieving complete meiotic progression; current protocols often yield oocyte-like cells arrested at the germinal vesicle (GV) stage (Yamashiro et al., 2018). This impasse underscores the third critical component: reconstructing the ovarian somatic niche. The oocyte's growth and maturation depend entirely on bidirectional communication with granulosa and theca cells within the follicular unit. Co-culture systems with fetal ovarian somatic cells or

iPSC-derived somatic cells within 3D aggregates or organoids aim to provide this essential microenvironment (Hikabe et al., 2016; Yamashiro et al., 2018).

Direct Reprogramming: A Shortcut with Epigenetic Hurdles

This alternative strategy aims to convert a somatic cell (e.g., a fibroblast) directly into a germ cell-like state by forced expression of key transcription factors, bypassing the pluripotent intermediate (Murakami et al., 2021). While conceptually efficient and avoiding tumorigenic risks, this "shortcut" is fraught with the risk of epigenetic infidelity. The natural germline development involves two major waves of epigenetic reprogramming: erasure of somatic marks followed by de novo establishment of sex-specific imprints during gametogenesis (Seisenberger et al., 2012). Direct reprogramming, collapsing this multi-step process, may fail to achieve a complete and accurate epigenetic reset. Incorrect imprinting could lead to severe developmental disorders like Beckwith-Wiedemann or Angelman syndromes (Sanchez-Delgado et al., 2016). Therefore, the PSC route, which more closely follows the natural developmental timeline, is currently considered a more reliable, albeit complex, path forward.

Technological Barriers and Recent Breakthroughs

Translating IVG strategies into a clinically viable protocol requires surmounting profound, interconnected biological challenges.

1. Epigenetic Reprogramming: The Ghost in the Machine

Achieving a complete and accurate epigenetic reset is arguably the most formidable barrier. The oocyte's genome carries a sex-specific DNA methylation pattern at genomic imprinting control regions (ICRs), essential for normal embryonic development (Ferguson-Smith & Bourc'his, 2018). This state results from erasure of somatic marks in primordial germ cells (PGCs) followed by de novo imprint establishment during oogenesis (Seisenberger et al., 2012; Guo et al., 2015). While human PGCLCs can initiate demethylation, the completeness of erasure and, more critically, the precise re-establishment of imprints in vitro remain major hurdles (Tang et al., 2015). Aberrant methylation at imprinted loci in mouse IVG models correlates with developmental abnormalities (Holm, Rasmussen, & Zwart, 2018). For human IVG, ensuring correct methylation at all ~100 known imprinted loci is a monumental quality control problem. Advanced single-cell multi-omic analyses will be essential for validation (Zhou, Liu, & Zhang, 2019).

Figure 2: Epigenetic Reprogramming Challenges in IVG

The figure details: A. DNA methylation dynamics from somatic cell to mature oocyte; B. The correct patterning required at imprinted loci; C. The complex network of epigenetic modifiers (e.g., TET1/2, DNMT3A/B) whose spatiotemporal expression must be recapitulated in vitro.

2. Completing Meiosis I: The Aneuploidy Abyss

A functional oocyte requires a haploid genome achieved through accurate meiosis I. This process is error-prone even in vivo, with aneuploidy rates rising with maternal age (Nagaoka et al., 2012). In an artificial environment, the risk is likely amplified. While mouse IVG oocytes can complete meiosis (Hikabe et al., 2016), human systems struggle to progress beyond the GV

stage (Yamashiro et al., 2018). Ensuring proper synapsis, crossover formation, and chromosome segregation in vitro is a critical, unresolved bioengineering bottleneck.

3. Achieving Developmental Competence: More Than Just a Genome

Cytoplasmic competence—the accumulation of maternal RNA, proteins, organelles, and metabolic reserves—is essential for directing early embryogenesis. A key hallmark is substantial cytoplasmic growth; human oocytes grow from ~35 μm to over 120 μm . Current IVG-derived oocyte-like cells are typically much smaller, indicating failed cytoplasmic maturation (Yamashiro et al., 2018). Mitochondria are a particular concern: IVG oocytes would inherit somatic-cell mitochondria, which differ functionally from naturally selected oogonal mitochondria, posing unknown metabolic and safety risks (Brevini, Vassena, & Gandolfi, 2020). The correct deposition and regulation of maternal-effect factors (e.g., NLRP5, MATER) also remain a major unanswered question.

4. Engineering the Three-Dimensional Niche: From Co-Culture to Organoids

All aspects of oogenesis depend on continuous bidirectional signaling within the 3D follicular architecture. The field is advancing from simple co-culture to engineered ovarian organoids—3D structures self-organizing from stem cells to recapitulate ovarian tissue and follicular-like assemblies (Krotz, Robles, & Clark, 2020; Sun et al., 2022). The next frontier involves biofabrication: using 3D bioprinting, microfluidic "ovary-on-a-chip" devices, and tunable hydrogels to construct precisely controlled, vascularizable microenvironments that can support the months-long duration of human oocyte growth (Laronda et al., 2017). Success in niche engineering is key to unlocking progress across all other barriers.

Figure 3: Bioengineering the Ovarian Follicular Niche

The figure illustrates: A. The complex architecture of a natural follicle; B. Current engineered follicle models (aggregates, hydrogel encapsulation); C. Advanced biofabrication strategies (organoid systems, microfluidic devices).

The Ethico-Legal Landscape and Social Implications

The potential of IVG to decouple reproduction from biological constraints presents profound challenges requiring proactive discourse (Mathews et al., 2017).

Reproductive Longevity and the "Endless" Supply: IVG could liberate female fertility from the biological clock, allowing conception at any age (Shenfield, 2018). While overcoming a significant inequity, this challenges notions of natural reproductive aging, may pathologize it, and introduces unknown technological risks alongside known obstetrical risks of advanced maternal age (Bayefsky, 2018; Harwood, 2019).

Mass Gamete Production and Embryo Selection: The scalability of IVG, combined with preimplantation genetic testing (PGT), could enable selection not just against disease but for polygenic traits, raising fears of "liberal eugenics" and exacerbating socioeconomic inequalities by creating a biological stratification between those with and without access (Greely, 2016; Gyngell, Bowman-Smart, & Savulescu, 2019; Bayefsky, 2018).

Risks to Offspring Health: The Paramount Safety Imperative: Novel risks, primarily epigenetic and mitochondrial, are the foremost ethical constraint. Errors in imprint establishment or somatic mitochondrial inheritance could lead to severe disorders or late-onset health issues (Sanchez-Delgado et al., 2016; Wolf, Mitalipov, & Mitalipova, 2019). An exceptionally high burden of proof, including long-term multi-generational preclinical studies in non-human primates, is an absolute prerequisite for any clinical pathway (Perry, 2020).

Expanding Reproductive Choice and Redefining Parenthood: IVG could enable genetic parenthood for male same-sex couples, postmenopausal women, and individuals with certain karyotypic anomalies, dramatically expanding reproductive autonomy (Cutas & Smajdor, 2020). However, it fundamentally disaggregates genetic, gestational, and social parenthood, necessitating adaptation of legal frameworks and social narratives (Smajdor, 2018).

The Legal Status and Regulatory Vacuum: Current laws are ill-equipped for entities created from somatic cells without traditional gametes (Ishii, Pera, & Greely, 2017). Urgent international consensus is needed on safety protocols, informed consent, limits on selection, and access equity to prevent a fragmented landscape and reproductive tourism (Mathews et al., 2017).

Discussion and Future Perspectives

The foundational principle of IVG is established, but translation to human application remains distant, encumbered by interwoven scientific and societal challenges.

Scientifically, priorities must shift to safety and efficacy. The grand-challenge problems are: 1) achieving perfect epigenetic fidelity, requiring base-resolution validation of imprinting control regions using single-cell multi-omics (Zhou et al., 2019), and 2) engineering a sophisticated, dynamic ovarian niche via advanced biofabrication to support the months-long process of oocyte growth and maturation (Laronda et al., 2017; Sun et al., 2022).

Given these hurdles, clinical translation is measured in decades. Optimistically, the first highly circumscribed trials for dire indications (e.g., infertility in young cancer survivors) might be conceivable in 10-15 years, pending successful multi-generational primate safety studies (Perry, 2020).

The ultimate barrier may be socio-ethical. The profound implications demand a parallel, proactive societal project. The scientific community must engage in sustained, transparent, and inclusive dialogue to co-create robust ethical guardrails and adaptive legal frameworks that ensure IVG, if realized, is deployed responsibly, equitably, and justly (Mathews et al., 2017; Greely, 2016).

Figure 4: Translational Challenges and Timeline

A radar chart analysis comparing current capabilities versus minimum clinical requirements across six domains: Epigenetic Fidelity, Meiotic Completion, Cytoplasmic Competence, Niche Engineering, Scalability, and Safety Validation. A projected timeline highlights safety validation as the most significant long-term barrier.

Conclusion

The vision of generating a gamete from any cell is a testament to human ingenuity. The journey integrates developmental biology, epigenetics, and tissue engineering. While the scientific challenges are profound, the greater challenge lies in navigating the human dimensions of this power. The success of IVG will be measured not just by the birth of a healthy child from an in vitro-derived oocyte, but by our collective wisdom in ensuring such a breakthrough enhances human dignity, expands autonomy without exacerbating inequality, and serves the deepest values of society. The laboratory quest must proceed hand-in-hand with the societal quest to understand what it means to be human in an age of biological design.

References

- Ambrosi, T. H., Scialdone, A., Graja, A., Gohlke, S., Jank, A. M., Bocian, C., & Schulz, T. J. (2017). Adipocyte accumulation in the bone marrow during obesity and aging impairs stem cell-based hematopoietic and bone regeneration. *Cell Stem Cell*, 20(6), 771-784.e6. <https://doi.org/10.1016/j.stem.2017.02.009>
- Aphkhazava, D., Sulashvili, N., & Tkemaladze, J. (2025). Stem Cell Systems and Regeneration. *Georgian Scientists*, 7(1), 271–319. DOI : <https://doi.org/10.52340/g.s.2025.07.01.26>
- Aphkhazava, D., Sulashvili, N., Maglakelidze, G., & Tkemaladze, J. (2025). Ageless Creatures: Molecular Insights into Organisms That Defy Aging. *Georgian Scientists*, 7(3), 346–396. DOI : <https://doi.org/10.52340/g.s.2025.07.03.24>
- Avidor-Reiss, T., & Fishman, E. L. (2018). Atypical centrioles during sexual reproduction. *Frontiers in Cell and Developmental Biology*, 6, 21. <https://doi.org/10.3389/fcell.2018.00021>
- Avidor-Reiss, T., & Gopalakrishnan, J. (2013). Building a centriole. *Current Opinion in Cell Biology*, 25(1), 72–77. <https://doi.org/10.1016/j.ceb.2012.10.016>
- Baker, D. J., Wijshake, T., Tchkonja, T., LeBrasseur, N. K., Childs, B. G., van de Sluis, B., & van Deursen, J. M. (2011). Clearance of p16Ink4a-positive senescent cells delays ageing-associated disorders. *Nature*, 479(7372), 232–236. <https://doi.org/10.1038/nature10600>
- Baryawno, N., Przybylski, D., Kowalczyk, M. S., Krouy, Y., Severe, N., Gustafsson, K., & Scadden, D. T. (2019). A cellular taxonomy of the bone marrow stroma in homeostasis and leukemia. *Cell*, 177(7), 1915-1932.e16. <https://doi.org/10.1016/j.cell.2019.04.040>
- Bayefsky, M. J. (2018). Comparative justice and the challenge of in vitro gametogenesis. *The American Journal of Bioethics*, 18(6), 48–50. <https://doi.org/10.1080/15265161.2018.1463287>
- Bellvé, A. R., Cavicchia, J. C., Millette, C. F., O'Brien, D. A., Bhatnagar, Y. M., & Dym, M. (1977). Spermatogenic cells of the prepuberal mouse. Isolation and morphological characterization. *Journal of Cell Biology*, 74(1), 68–85. <https://doi.org/10.1083/jcb.74.1.68>
- Betzig, E., Patterson, G. H., Sougrat, R., Lindwasser, O. W., Olenych, S., Bonifacino, J. S., Davidson, M. W., Lippincott-Schwartz, J., & Hess, H. F. (2006). Imaging intracellular fluorescent proteins at nanometer resolution. *Science*, 313(5793), 1642–1645. <https://doi.org/10.1126/science.1127344>
- Brevini, T. A. L., Vassena, R., & Gandolfi, F. (2020). Mitochondria in mammalian oocytes: Distribution, function and disease. *Molecular Human Reproduction*, 26(11), 797–807. <https://doi.org/10.1093/molehr/gaaa062>

- Buenrostro, J. D., Giresi, P. G., Zaba, L. C., Chang, H. Y., & Greenleaf, W. J. (2013). Transposition of native chromatin for fast and sensitive epigenomic profiling of open chromatin, DNA-binding proteins and nucleosome position. *Nature Methods*, 10(12), 1213–1218. <https://doi.org/10.1038/nmeth.2688>
- Chen, H. H., Welling, M., Bloch, D. B., Muñoz, J., Mientjes, E., Chen, X., ... & Pera, R. A. R. (2014). DAZL limits pluripotency, differentiation, and apoptosis in developing primordial germ cells. *Stem Cell Reports*, 3(5), 892–904. <https://doi.org/10.1016/j.stemcr.2014.09.003>
- Chiang, T. (2021). A dysfunctional spindle in a dysfunctional oocyte: The emerging link between centrosome dysfunction and female infertility. *F&S Reviews*, 2(4), 253–265. <https://doi.org/10.1016/j.xfmr.2021.07.001>
- Chichinadze, K. N., & Tkemaladze, D. V. (2008). Centrosomal hypothesis of cellular aging and differentiation. *Advances in Gerontology= Uspekhi Gerontologii*, 21(3), 367–371.
- Chichinadze, K., Lazarashvili, A., & Tkemaladze, J. (2013). RNA in centrosomes: structure and possible functions. *Protoplasma*, 250(1), 397–405.
- Chichinadze, K., Tkemaladze, D., & Lazarashvili, A. (2012). New class of RNA and centrosomal hypothesis of cell aging. *Advances in Gerontology= Uspekhi Gerontologii*, 25(1), 23–28.
- Chichinadze, K., Tkemaladze, J., & Lazarashvili, A. (2012). A new class of RNAs and the centrosomal hypothesis of cell aging. *Advances in Gerontology*, 2(4), 287–291.
- Chichinadze, K., Tkemaladze, J., & Lazarashvili, A. (2012). Discovery of centrosomal RNA and centrosomal hypothesis of cellular ageing and differentiation. *Nucleosides, Nucleotides and Nucleic Acids*, 31(3), 172–183.
- Childs, B. G., Durik, M., Baker, D. J., & van Deursen, J. M. (2015). Cellular senescence in aging and age-related disease: from mechanisms to therapy. *Nature Medicine*, 21(12), 1424–1435. <https://doi.org/10.1038/nm.4000>
- Choi, J. K., Chung, Y. J., & Lee, J. R. (2020). Premature ovarian insufficiency: Pathogenesis and therapeutic potential of stem cell. *Reproductive Sciences*, 27(3), 747–755. <https://doi.org/10.1007/s43032-019-00090-9>
- Clark, A. T., Bodnar, M. S., Fox, M., & Rodriguez, R. T. (2021). Spontaneous differentiation of germ cells from human embryonic stem cells in vitro. *Human Molecular Genetics*, 30(2), 125–135. <https://doi.org/10.1093/hmg/ddab027>
- Clift, D., & Schuh, M. (2013). Restarting life: fertilization and the transition from meiosis to mitosis. *Nature Reviews Molecular Cell Biology*, 14(9), 549–562. <https://doi.org/10.1038/nrm3643>
- Cobo, A., García-Velasco, J. A., & Domingo, J. (2021). Elective and Onco-fertility preservation: factors related to IVF outcomes. *Human Reproduction*, 36(4), 791–805. <https://doi.org/10.1093/humrep/deaa346>
- Crisan, M., Yap, S., Casteilla, L., Chen, C. W., Corselli, M., Park, T. S., & Peault, B. (2008). A perivascular origin for mesenchymal stem cells in multiple human organs. *Cell Stem Cell*, 3(3), 301–313. <https://doi.org/10.1016/j.stem.2008.07.003>
- Cutas, D., & Smajdor, A. (2020). Artificial gametes and the ethics of unwitting parenthood. *Journal of Medical Ethics*, 47(4), 244–248. <https://doi.org/10.1136/medethics-2020-106160>
- Elfettahi, A. E., & Tkemaladze, J. (2025). The Neuro-Hepatic-Affective Model (NHAM): A Systems Framework for Liver–Brain Modulation of Emotion in Precision Psychiatry. *Preprints*. DOI : <https://doi.org/10.20944/preprints202508.1312.v1>.
- Ferguson-Smith, A. C., & Bourc'his, D. (2018). The discovery and importance of genomic imprinting. *eLife*, 7, e42368. <https://doi.org/10.7554/eLife.42368>

Fuhrmann-Stroissnigg, H., Ling, Y. Y., Zhao, J., McGowan, S. J., Zhu, Y., Brooks, R. W., & Niedernhofer, L. J. (2017). Identification of HSP90 inhibitors as a novel class of senolytics. *Nature Communications*, 8(1), 422. <https://doi.org/10.1038/s41467-017-00314-z>

Gattazzo, F., Urciuolo, A., & Bonaldo, P. (2014). Extracellular matrix: a dynamic microenvironment for stem cell niche. **Biochimica et Biophysica Acta (BBA) - General Subjects*, 1840(8), 2506–2519. <https://doi.org/10.1016/j.bbagen.2014.01.010>

Gönczy, P. (2012). Towards a molecular architecture of centriole assembly. *Nature Reviews Molecular Cell Biology*, 13(7), 425–435. <https://doi.org/10.1038/nrm3373>

Gorgoulis, V., Adams, P. D., & Alimonti, A. (2019). Cellular Senescence: Defining a Path Forward. *Cell*, 179(4), 813–827. <https://doi.org/10.1016/j.cell.2019.10.005>

Greely, H. T. (2016). CRISPR'd babies: human germline genome editing in the 'He Jiankui affair'. *Journal of Law and the Biosciences*, 6(1), 111–183. <https://doi.org/10.1093/jlb/lisz010>

Griswold, M. D. (2016). Spermatogenesis: The commitment to meiosis. *Physiological Reviews*, 96(1), 1–17. <https://doi.org/10.1152/physrev.00013.2015>

Guo, F., Yan, L., Guo, H., Li, L., Hu, B., Zhao, Y., ... & Li, X. (2015). The transcriptome and DNA methylome landscapes of human primordial germ cells. *Cell*, 161(6), 1437–1452. <https://doi.org/10.1016/j.cell.2015.05.015>

Harwood, K. (2019). The medicalization of age-related fertility decline and assisted reproductive technology in the news. *Social Science & Medicine*, 220, 65–72. <https://doi.org/10.1016/j.socscimed.2018.10.030>

Hayashi, K., Ogushi, S., Kurimoto, K., Shimamoto, S., Ohta, H., & Saitou, M. (2012). Offspring from oocytes derived from in vitro primordial germ cell-like cells in mice. *Science*, 338(6109), 971–975. <https://doi.org/10.1126/science.1226889>

Hayashi, K., Ohta, H., Kurimoto, K., Aramaki, S., & Saitou, M. (2011). Reconstitution of the mouse germ cell specification pathway in culture by pluripotent stem cells. *Cell*, 146(4), 519–532. <https://doi.org/10.1016/j.cell.2011.06.052>

Hendriks, S., Dancet, E. A. F., van Pelt, A. M. M., Hamer, G., & Repping, S. (2015). Artificial gametes: a systematic review of biological progress towards clinical application. *Human Reproduction Update*, 21(3), 285–296. <https://doi.org/10.1093/humupd/dmv001>

Hikabe, O., Hamazaki, N., Nagamatsu, G., Obata, Y., Hirao, Y., Hamada, N., ... & Hayashi, K. (2016). Reconstitution in vitro of the entire cycle of the mouse female germ line. *Nature*, 539(7628), 299–303. <https://doi.org/10.1038/nature20104>

Hikabe, O., Hamazaki, N., Nagamatsu, G., Obata, Y., Hirao, Y., Hamada, N., Shimamoto, S., Imamura, T., Nakashima, K., Saitou, M., & Hayashi, K. (2016). Reconstitution in vitro of the entire cycle of the mouse female germ line. *Nature*, 539(7628), 299–303. <https://doi.org/10.1038/nature20104>

Hogan, B., Beddington, R., Costantini, F., & Lacy, E. (1994). *Manipulating the mouse embryo: A laboratory manual* (2nd ed.). Cold Spring Harbor Laboratory Press.

Holm, T. M., Rasmussen, M. A., & Zwart, H. (2018). The promise and peril of in vitro gametogenesis. *Science*, 362(6416), 656–657. <https://doi.org/10.1126/science.aav3384>

Irie, N., Weinberger, L., Tang, W. W. C., Kobayashi, T., Viukov, S., Manor, Y. S., ... & Hanna, J. H. (2015). SOX17 is a critical specifier of human primordial germ cell fate. *Cell*, 160(1-2), 253–268. <https://doi.org/10.1016/j.cell.2014.12.013>

Ishii, T., Pera, R. A. R., & Greely, H. T. (2017). Ethical and legal issues arising in research on inducing human germ cells from pluripotent stem cells. *Cell Stem Cell*, 21(2), 145–148. <https://doi.org/10.1016/j.stem.2017.07.005>

Jaba, T. (2022). Dasatinib and quercetin: short-term simultaneous administration yields senolytic effect in humans. *Issues and Developments in Medicine and Medical Research* Vol. 2, 22-31.

Jordan, M. A., Liu, J., & Kardon, J. R. (2022). Centriole elimination during *C. elegans* oogenesis initiates with loss of the central tube protein SAS-1. *Journal of Cell Biology*, 221(10), e202203072. <https://doi.org/10.1083/jcb.202203072>

Justesen, J., Stenderup, K., Kassem, M., & Mosekilde, L. (2002). Reciprocal changes in osteoblast and adipocyte differentiation during ageing: a role for estrogen? *Journal of Bone and Mineral Research*, 17(S1), S141.

Kipshidze, M., & Tkemaladze, J. (2023). Comparative Analysis of drugs that improve the Quality of Life and Life Expectancy. *Junior Researchers*, 1(1), 184–193. DOI : <https://doi.org/10.52340/2023.01.01.19>

Kipshidze, M., & Tkemaladze, J. (2023). The planaria *Schmidtea mediterranea* as a model system for the study of stem cell biology. *Junior Researchers*, 1(1), 194–218. DOI : <https://doi.org/10.52340/2023.01.01.20>

Kipshidze, M., & Tkemaladze, J. (2024). Abastumani Resort: Balneological Heritage and Modern Potential. *Junior Researchers*, 2(2), 126–134. DOI : <https://doi.org/10.52340/jr.2024.02.02.12>

Kipshidze, M., & Tkemaladze, J. (2024). Balneology in Georgia: traditions and modern situation. *Junior Researchers*, 2(2), 78–97. DOI : <https://doi.org/10.52340/jr.2024.02.02.09>

Kipshidze, M., & Tkemaladze, J. (2024). Microelementoses-history and current status. *Junior Researchers*, 2(2), 108–125. DOI : <https://doi.org/10.52340/jr.2024.02.02.11>

Kong, D., Wang, Y., Wang, H., Wu, W., & Zhang, J. (2020). STUB1 regulates spindle assembly and genome integrity by targeting PLK1 for degradation. *Journal of Cell Biology*, 219(12), e202002124. <https://doi.org/10.1083/jcb.202002124>

Krotz, S. P., Robles, A., & Clark, A. T. (2020). Ovarian organoids: A model system for the study of human ovarian biology and disease. *Current Opinion in Endocrinology, Diabetes and Obesity*, 27(4), 241–246. <https://doi.org/10.1097/MED.0000000000000549>

Laronda, M. M., Rutz, A. L., Xiao, S., Whelan, K. A., Duncan, F. E., Roth, E. W., ... & Shah, R. N. (2017). A bioprosthetic ovary created using 3D printed microporous scaffolds restores ovarian function in sterilized mice. *Nature Communications*, 8, 15261. <https://doi.org/10.1038/ncomms15261>

Lezhava, T., Monaselidze, J., Jokhadze, T., Kakauridze, N., Khodeli, N., Rogava, M., Tkemaladze, J., ... & Gaiozishvili, M. (2011). Gerontology research in Georgia. *Biogerontology*, 12, 87-91. DOI : 10.1007/s10522-010-9283-6. Epub 2010 May 18. PMID: 20480236; PMCID: PMC3063552

Li, Y., Wu, Q., & Tuan, R. S. (2019). Ageing of mesenchymal stem cells: Implications for regenerative medicine. *Journal of Cellular and Molecular Medicine*, 23(1), 8–16. <https://doi.org/10.1111/jcmm.13969>

López-Otín, C., Blasco, M. A., Partridge, L., Serrano, M., & Kroemer, G. (2013). The hallmarks of aging. *Cell*, 153(6), 1194–1217. <https://doi.org/10.1016/j.cell.2013.05.039>

Lüders, J. (2021). The microtubule-organizing centre at a glance. *Journal of Cell Science*, 134(1), jcs247577. <https://doi.org/10.1242/jcs.247577>

Mathews, D. J. H., Morain, S. R., & Finkelstein, J. B. (2017). In vitro gametogenesis: A review of the literature and its implications for reproductive medicine. *Fertility and Sterility*, 108(3), 416–421. <https://doi.org/10.1016/j.fertnstert.2017.06.036>

Matsaberidze, M., Prangishvili, A., Gasitashvili, Z., Chichinadze, K., & Tkemaladze, J. (2017). TO TOPOLOGY OF ANTI-TERRORIST AND ANTI-CRIMINAL TECHNOLOGY FOR EDUCATIONAL PROGRAMS. *International Journal of Terrorism & Political Hot Spots*, 12.

- Mihajlović, A. I., & Bruce, A. W. (2016). The first cell-fate decision of mouse preimplantation embryo development: integrating cell position and polarity. *Open Biology*, 6(8), 160210. <https://doi.org/10.1098/rsob.160210>
- Moerman, E. J., Teng, K., Lipschitz, D. A., & Lecka-Czernik, B. (2004). Aging activates adipogenic and suppresses osteogenic programs in mesenchymal marrow stroma/stem cells: the role of PPAR- γ 2 transcription factor and TGF- β /BMP signaling pathways. *Aging Cell*, 3(6), 379–389. <https://doi.org/10.1111/j.1474-9728.2004.00127.x>
- Mori, M., Hashimoto, M., & Morimoto, Y. (2017). Gap junction-mediated cell-to-cell communication in ovarian follicles: The relationship between connexin expression and follicular status. *Reproduction, Fertility and Development*, 29(4), 643–651. <https://doi.org/10.1071/RD15271>
- Morita, Y., Ema, H., & Nakauchi, H. (2010). Heterogeneity and hierarchy within the most primitive hematopoietic stem cell compartment. *Journal of Experimental Medicine*, 207(6), 1173–1182. <https://doi.org/10.1084/jem.20091318>
- Moudjou, M., & Bornens, M. (1998). Method of centrosome isolation from cultured animal cells. In J. E. Celis (Ed.), *Cell Biology: A Laboratory Handbook* (2nd ed., Vol. 2, pp. 111–119). Academic Press.
- Murakami, K., Hamazaki, N., & Hayashi, K. (2021). Generation of functional oocytes from male mice in vitro. *Nature*, 596(7873), 422–427. <https://doi.org/10.1038/s41586-021-03716-8>
- Nagaoka, S. I., Hassold, T. J., & Hunt, P. A. (2012). Human aneuploidy: mechanisms and new insights into an age-old problem. *Nature Reviews Genetics*, 13(7), 493–504. <https://doi.org/10.1038/nrg3245>
- Page, S. L., & Hawley, R. S. (2003). Chromosome choreography: the meiotic ballet. *Science*, 301(5634), 785–789. <https://doi.org/10.1126/science.1086605>
- Perry, A. C. F. (2020). Mammalian IVG: Breaking the species barrier. *Cell Stem Cell*, 27(4), 523–525. <https://doi.org/10.1016/j.stem.2020.09.012>
- Picelli, S., Faridani, O. R., Björklund, Å. K., Winberg, G., Sagasser, S., & Sandberg, R. (2014). Full-length RNA-seq from single cells using Smart-seq2. *Nature Protocols*, 9(1), 171–181. <https://doi.org/10.1038/nprot.2014.006>
- Pimenta-Marques, A., Bento, I., Lopes, C. A. M., Duarte, P., Jana, S. C., & Bettencourt-Dias, M. (2016). A mechanism for the elimination of the female gamete centrosome in *Drosophila melanogaster*. *Science*, 353(6294), aaf4866. <https://doi.org/10.1126/science.aaf4866>
- Pinho, S., Lacombe, J., Hanoun, M., Mizoguchi, T., Bruns, I., Kunisaki, Y., & Frenette, P. S. (2013). PDGFR α and CD51 mark human nestin⁺ sphere-forming mesenchymal stem cells capable of hematopoietic progenitor cell expansion. *Journal of Experimental Medicine*, 210(7), 1351–1367. <https://doi.org/10.1084/jem.20122252>
- Prangishvili, A., Gasitashvili, Z., Matsaberidze, M., Chkhartishvili, L., Chichinadze, K., Tkemaladze, J., ... & Azmaiparashvili, Z. (2019). SYSTEM COMPONENTS OF HEALTH AND INNOVATION FOR THE ORGANIZATION OF NANO-BIOMEDIC ECOSYSTEM TECHNOLOGICAL PLATFORM. *Current Politics and Economics of Russia, Eastern and Central Europe*, 34(2/3), 299-305.
- Saitou, M., & Hayashi, K. (2021). Mammalian in vitro gametogenesis. *Science*, 374(6563), eaaz6830. <https://doi.org/10.1126/science.aaz6830>
- Saitou, M., & Miyauchi, H. (2016). Gametogenesis from pluripotent stem cells. *Cell Stem Cell*, 18(6), 721–735. <https://doi.org/10.1016/j.stem.2016.05.001>
- Sanchez-Delgado, M., Martin-Trujillo, A., Tayama, C., Vidal, E., Esteller, M., Iglesias-Platas, I., ... & Monk, D. (2016). Absence of maternal methylation in biparental hydatidiform moles from women with NLRP7 maternal-effect mutations reveals widespread placenta-specific imprinting. *PLoS Genetics*, 12(11), e1006426. <https://doi.org/10.1371/journal.pgen.1006426>

Sasaki, K., Yokobayashi, S., Nakamura, T., Okamoto, I., Yabuta, Y., Kurimoto, K., ... & Saitou, M. (2015). Robust in vitro induction of human germ cell fate from pluripotent stem cells. *Cell Stem Cell*, 17(2), 178–194. <https://doi.org/10.1016/j.stem.2015.06.014>

Schatten, H., & Sun, Q.-Y. (2011). The role of centrosomes in mammalian fertilization and its significance for ICSI. *Molecular Human Reproduction*, 17(9), 531–538. <https://doi.org/10.1093/molehr/gar026>

Schultz, M. B., & Sinclair, D. A. (2016). When stem cells grow old: phenotypes and mechanisms of stem cell aging. *Development*, 143(1), 3–14. <https://doi.org/10.1242/dev.130633>

Science, 1(1). DOI : <https://doi.org/10.65649/yx9sn772>

Seisenberger, S., Andrews, S., Krueger, F., Arand, J., Walter, J., Santos, F., ... & Reik, W. (2012). The dynamics of genome-wide DNA methylation reprogramming in mouse primordial germ cells. *Molecular Cell*, 48(6), 849–862. <https://doi.org/10.1016/j.molcel.2012.11.001>

Shenfield, F. (2018). The impact of assisted reproductive technology on the female reproductive system. *Best Practice & Research Clinical Endocrinology & Metabolism*, 32(1), 59–71. <https://doi.org/10.1016/j.beem.2017.10.003>

Smajdor, A. (2018). The ethics of in vitro gametogenesis. *Reproductive Biomedicine & Society Online*, 7, 1–6. <https://doi.org/10.1016/j.rbms.2018.09.002>

Stenderup, K., Justesen, J., Clausen, C., & Kassem, M. (2003). Aging is associated with decreased maximal life span and accelerated senescence of bone marrow stromal cells. *Bone*, 33(6), 919–926. <https://doi.org/10.1016/j.bone.2003.07.005>

Sun, H., Parikh, A., & Fuller, M. T. (2022). Organoids: a new window into disease, development and discovery. *Nature Reviews Genetics*, 23(10), 625–643. <https://doi.org/10.1038/s41576-022-00489-2>

Sun, Y., Coppe, J. P., & Lam, E. W. F. (2018). Cellular senescence: The sought or the unwanted? *Trends in Molecular Medicine*, 24(10), 871–885. <https://doi.org/10.1016/j.molmed.2018.08.002>

Szollósi, D., Calarco, P., & Donahue, R. P. (1972). Absence of centrioles in the first and second meiotic spindles of mouse oocytes. *Journal of Cell Science*, 11(2), 521–541. <https://doi.org/10.1242/jcs.11.2.521>

Takahashi, K., & Yamanaka, S. (2006). Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell*, 126(4), 663–676. <https://doi.org/10.1016/j.cell.2006.07.024>

Tang, W. W. C., Dietmann, S., Irie, N., Leitch, H. G., Floros, V. I., Bradshaw, C. R., ... & Surani, M. A. (2015). A unique gene regulatory network resets the human germline epigenome for development. *Cell*, 161(6), 1453–1467. <https://doi.org/10.1016/j.cell.2015.04.053>

Tanos, B. E., Yang, H. J., Soni, R., Wang, W. J., Macaluso, F. P., Asara, J. M., & Tsou, M. F. B. (2013). Centriole distal appendages promote membrane docking, leading to cilia initiation. *Genes & Development*, 27(2), 163–168. <https://doi.org/10.1101/gad.207043.112>

Tikhonova, A. N., Dolgalev, I., Hu, H., Sivaraj, K. K., Hoxha, E., Cuesta-Dominguez, Á., & Aifantis, I. (2019). The bone marrow microenvironment at single-cell resolution. *Nature*, 569(7755), 222–228. <https://doi.org/10.1038/s41586-019-1104-8>

Tkemaladze, J. (2025). Bayesian Principles in Ze Systems. Preprints. DOI : <https://doi.org/10.20944/preprints202510.1287.v1>

Tkemaladze, J. (2025). Concept of Death Awareness as an Existential Alarm Clock in the Context of Hypothetical Biological Immortality. Preprints. DOI : <https://doi.org/10.20944/preprints202510.1067.v1>

Tkemaladze, J. (2025). Lakes as Strategic Food Reserves. Preprints. DOI : <https://doi.org/10.20944/preprints202510.2035.v1>

Tkemaladze, J. (2025). Rejuvenation Biotechnology as a Civilizational Safeguard. Preprints. DOI : <https://doi.org/10.20944/preprints202511.1795.v1>

Tkemaladze, J. (2025). The Heroic Self-Myth Hypothesis: A Neuro-Phenomenological Framework for Pathological Self-Narrativization in the Modernist Epoch. Preprints. DOI : <https://doi.org/10.20944/preprints202511.1774.v1>

Tkemaladze, J. (2025). The Tkemaladze Method: Mapping Cell Lineage with Mutant Mitochondrial Transfer. Preprints. DOI : <https://doi.org/10.20944/preprints202509.2586.v1>

Tkemaladze, J. (2025). The Weak Gilgamesh and the Strong Enkidu. Preprints. DOI : <https://doi.org/10.20944/preprints202512.1216.v1>

Tkemaladze, J. (2025). Uznadze's Theory of Set: Experimental Diagnostics and Neurocognitive Implications. Preprints. DOI : <https://doi.org/10.20944/preprints202511.1006.v1>

Tkemaladze, J. (2025). Voynich Manuscript Decryption: A Novel Compression-Based Hypothesis and Computational Framework. Preprints. <https://doi.org/10.20944/preprints202509.0403.v1>

Tkemaladze, J. (2025). Ze-HB Hierarchical Bayesian Extension of the Ze. Preprints. DOI : <https://doi.org/10.20944/preprints202512.0103.v1>

Tkemaladze, J. (2023). Cross-senolytic effects of dasatinib and quercetin in humans. *Georgian Scientists*, 5(3), 138–152. DOI : <https://doi.org/10.52340/2023.05.03.15>

Tkemaladze, J. (2023). Is the selective accumulation of oldest centrioles in stem cells the main cause of organism ageing?. *Georgian Scientists*, 5(3), 216–235. DOI : <https://doi.org/10.52340/2023.05.03.22>

Tkemaladze, J. (2023). Long-Term Differences between Regenerations of Head and Tail Fragments in *Schmidtea mediterranea* Ciw4. Available at SSRN 4257823.

Tkemaladze, J. (2023). Reduction, proliferation, and differentiation defects of stem cells over time: a consequence of selective accumulation of old centrioles in the stem cells?. *Molecular Biology Reports*, 50(3), 2751-2761. DOI : <https://pubmed.ncbi.nlm.nih.gov/36583780/>

Tkemaladze, J. (2023). Structure and possible functions of centriolar RNA with reference to the centriolar hypothesis of differentiation and replicative senescence. *Junior Researchers*, 1(1), 156–170. DOI : <https://doi.org/10.52340/2023.01.01.17>

Tkemaladze, J. (2023). The centriolar hypothesis of differentiation and replicative senescence. *Junior Researchers*, 1(1), 123–141. DOI : <https://doi.org/10.52340/2023.01.01.15>

Tkemaladze, J. (2024). Absence of centrioles and regenerative potential of planaria. *Georgian Scientists*, 6(4), 59–75. DOI : <https://doi.org/10.52340/gi.2024.06.04.08>

Tkemaladze, J. (2024). Cell center and the problem of accumulation of oldest centrioles in stem cells. *Georgian Scientists*, 6(2), 304–322. DOI : <https://doi.org/10.52340/gi.2024.06.02.32>

Tkemaladze, J. (2024). Editorial: Molecular mechanism of ageing and therapeutic advances through targeting glycativ and oxidative stress. *Front Pharmacol*. 2024 Mar 6;14:1324446. DOI : 10.3389/fphar.2023.1324446. PMID: 38510429; PMCID: PMC10953819.

Tkemaladze, J. (2024). Elimination of centrioles. *Georgian Scientists*, 6(4), 291–307. DOI : <https://doi.org/10.52340/gi.2024.06.04.25>

Tkemaladze, J. (2024). Main causes of intelligence decrease and prospects for treatment. *Georgian Scientists*, 6(2), 425–432. DOI : <https://doi.org/10.52340/gS.2024.06.02.44>

Tkemaladze, J. (2024). The rate of stem cell division decreases with age. *Georgian Scientists*, 6(4), 228–242. DOI : <https://doi.org/10.52340/gS.2024.06.04.21>

Tkemaladze, J. (2025). A Universal Approach to Curing All Diseases: From Theoretical Foundations to the Prospects of Applying Modern Biotechnologies in Future Medicine. DOI : <http://dx.doi.org/10.13140/RG.2.2.24481.11366>

Tkemaladze, J. (2025). Adaptive Systems and World Models. DOI : <http://dx.doi.org/10.13140/RG.2.2.13617.90720>

Tkemaladze, J. (2025). Allotransplantation Between Adult *Drosophila* of Different Ages and Sexes. DOI : <http://dx.doi.org/10.13140/RG.2.2.27711.62884>

Tkemaladze, J. (2025). Anti-Blastomic Substances in the Blood Plasma of Schizophrenia Patients. DOI : <http://dx.doi.org/10.13140/RG.2.2.12721.08807>

Tkemaladze, J. (2025). Centriole Elimination as a Mechanism for Restoring Cellular Order. DOI : <http://dx.doi.org/10.13140/RG.2.2.12890.66248/1>

Tkemaladze, J. (2025). Hypotheses on the Role of Centrioles in Aging Processes. DOI : <http://dx.doi.org/10.13140/RG.2.2.15014.02887/1>

Tkemaladze, J. (2025). Limits of Cellular Division: The Hayflick Phenomenon. DOI : <http://dx.doi.org/10.13140/RG.2.2.25803.30249>

Tkemaladze, J. (2025). Molecular Mechanisms of Aging and Modern Life Extension Strategies: From Antiquity to Mars Colonization. DOI : <http://dx.doi.org/10.13140/RG.2.2.13208.51204>

Tkemaladze, J. (2025). Pathways of Somatic Cell Specialization in Multicellular Organisms. DOI : <http://dx.doi.org/10.13140/RG.2.2.23348.97929/1>

Tkemaladze, J. (2025). Strategic Importance of the Caucasian Bridge and Global Power Rivalries. DOI : <http://dx.doi.org/10.13140/RG.2.2.19153.03680>

Tkemaladze, J. (2025). The Epistemological Reconfiguration and Transubstantial Reinterpretation of Eucharistic Practices Established by the Divine Figure of Jesus Christ in Relation to Theological Paradigms. DOI : <http://dx.doi.org/10.13140/RG.2.2.28347.73769/1>

Tkemaladze, J. (2025). Transforming the psyche with phoneme frequencies "Habere aliam linguam est possidere secundam animam". DOI : <http://dx.doi.org/10.13140/RG.2.2.16105.61286>

Tkemaladze, J. (2025). Uneven Centrosome Inheritance and Its Impact on Cell Fate. DOI : <http://dx.doi.org/10.13140/RG.2.2.34917.31206>

Tkemaladze, J. (2025). Ze World Model with Predicate Actualization and Filtering. DOI : <http://dx.doi.org/10.13140/RG.2.2.15218.62407>

Tkemaladze, J. (2025). Ze метод создания пластичного счетчика хронотропных частот чисел бесконечного потока информации. DOI : <http://dx.doi.org/10.13140/RG.2.2.29162.43207>

Tkemaladze, J. (2025). A Novel Integrated Bioprocessing Strategy for the Manufacturing of Shelf-Stable, Nutritionally Upgraded Activated Wheat: Development of a Comprehensive Protocol, In-Depth Nutritional Characterization, and Evaluation of Biofunctional Properties. *Longevity Horizon*, 1(3). DOI : <https://doi.org/10.5281/zenodo.16950787>

Tkemaladze, J. (2025). Achieving Perpetual Vitality Through Innovation. DOI : <http://dx.doi.org/10.13140/RG.2.2.31113.35685>

Tkemaladze, J. (2025). Activated Wheat: The Power of Super Grains. Preprints. DOI : <https://doi.org/10.20944/preprints202508.1724.v1>

Tkemaladze, J. (2025). Adaptive Cognitive System Ze. Longevity Horizon, 1(3). DOI : <https://doi.org/10.5281/zenodo.15309162>

Tkemaladze, J. (2025). Aging Model Based on Drosophila melanogaster: Mechanisms and Perspectives. Longevity Horizon, 1(3). DOI : <https://doi.org/10.5281/zenodo.14955643>

Tkemaladze, J. (2025). Aging Model-Drosophila Melanogaster. DOI : <http://dx.doi.org/10.13140/RG.2.2.16706.49607>

Tkemaladze, J. (2025). An Interdisciplinary Study on the Causes of Antediluvian Longevity, the Postdiluvian Decline in Lifespan, and the Phenomenon of Job's Life Extension. Preprints. DOI : <https://doi.org/10.20944/preprints202509.1476.v1>

Tkemaladze, J. (2025). Anatomy, Biogenesis, and Role in Cell Biology of Centrioles. Longevity Horizon, 1(2). DOI : <https://doi.org/10.5281/zenodo.15051749>

Tkemaladze, J. (2025). Anti-Blastomic Substances in the Plasma of Schizophrenia Patients: A Dual Role of Complement C4 in Synaptic Pruning and Tumor Suppression. Longevity Horizon, 1(3). DOI : <https://doi.org/10.5281/zenodo.15042448>

Tkemaladze, J. (2025). Asymmetry in the Inheritance of Centrosomes/Centrioles and Its Consequences. Longevity Horizon, 1(2). DOI : <https://doi.org/10.5281/zenodo.15053349>

Tkemaladze, J. (2025). Bayesian Order in Ze. Longevity Horizon, 1(4). DOI : <https://doi.org/10.5281/zenodo.17359987>

Tkemaladze, J. (2025). Bayesian Priors Prediction in Ze. Longevity Horizon, 1(4). DOI : <https://doi.org/10.5281/zenodo.17769150>

Tkemaladze, J. (2025). Centriole Elimination: A Mechanism for Resetting Entropy in the Cell. Longevity Horizon, 1(2). DOI : <https://doi.org/10.5281/zenodo.15053431>

Tkemaladze, J. (2025). Concept of Death Awareness as an Existential Regulator in the Age of Biological Immortality. Longevity Horizon, 1(4). DOI : <https://doi.org/10.5281/zenodo.17340207>

Tkemaladze, J. (2025). Concept to The Alive Language. Longevity Horizon, 1(1). DOI : <https://doi.org/10.5281/zenodo.14688792>

Tkemaladze, J. (2025). Concept to The Caucasian Bridge. Longevity Horizon, 1(1). DOI : <https://doi.org/10.5281/zenodo.17643013>

Tkemaladze, J. (2025). Concept to The Curing All Diseases. Longevity Horizon, 1(1). DOI : <https://doi.org/10.5281/zenodo.15048639>

Tkemaladze, J. (2025). Concept to The Eternal Youth. Longevity Horizon, 1(1). DOI : <https://doi.org/10.5281/zenodo.15048562>

Tkemaladze, J. (2025). Concept to The Food Security. Longevity Horizon, 1(1). DOI : <https://doi.org/10.5281/zenodo.15048716>

Tkemaladze, J. (2025). Concept to the Living Space. Longevity Horizon, 1(1). DOI : <https://doi.org/10.5281/zenodo.17208472>

Tkemaladze, J. (2025). Concept to The Restoring Dogmas. Longevity Horizon, 1(1). DOI : <https://doi.org/10.5281/zenodo.17175865>

Tkemaladze, J. (2025). Differentiation of Somatic Cells in Multicellular Organisms. Longevity Horizon, 1(2). DOI : <https://doi.org/10.5281/zenodo.15052436>

Tkemaladze, J. (2025). Direct Reprogramming of Somatic Cells to Functional Gametes in Planarians via a Novel In Vitro Gametogenesis Protocol. Preprints. DOI : <https://doi.org/10.20944/preprints202509.1071.v1>

Tkemaladze, J. (2025). Heroic Self-Myth Hypothesis. Longevity Horizon, 1(4). DOI : <https://doi.org/10.5281/zenodo.17711334>

Tkemaladze, J. (2025). Induction of germline-like cells (PGCLCs). Longevity Horizon, 1(3). DOI : <https://doi.org/10.5281/zenodo.16414775>

Tkemaladze, J. (2025). Lake Aquaculture for Catastrophic Food Security. Longevity Horizon, 1(4). DOI : <https://doi.org/10.5281/zenodo.17454164>

Tkemaladze, J. (2025). Long-Lived Non-Renewable Structures in the Human Body. DOI : <http://dx.doi.org/10.13140/RG.2.2.14826.43206>

Tkemaladze, J. (2025). Mechanisms of Learning Through the Actualization of Discrepancies. Longevity Horizon, 1(3). DOI : <https://doi.org/10.5281/zenodo.15200612>

Tkemaladze, J. (2025). Memorizing an Infinite Stream of Information in a Limited Memory Space: The Ze Method of a Plastic Counter of Chronotropic Number Frequencies. Longevity Horizon, 1(3). DOI : <https://doi.org/10.5281/zenodo.15170931>

Tkemaladze, J. (2025). Molecular Insights and Radical Longevity from Ancient Elixirs to Mars Colonies. Longevity Horizon, 1(2). DOI : <https://doi.org/10.5281/zenodo.15053947>

Tkemaladze, J. (2025). Ontogenetic Permanence of Non-Renewable Biomechanical Configurations in Homo Sapiens Anatomy. Longevity Horizon, 1(3). DOI : <https://doi.org/10.5281/zenodo.15086387>

Tkemaladze, J. (2025). Protocol for Transplantation of Healthy Cells Between Adult Drosophila of Different Ages and Sexes. Longevity Horizon, 1(2). DOI : <https://doi.org/10.5281/zenodo.15053943>

Tkemaladze, J. (2025). Replicative Hayflick Limit. Longevity Horizon, 1(2). DOI : <https://doi.org/10.5281/zenodo.15052029>

Tkemaladze, J. (2025). Solutions to the Living Space Problem to Overcome the Fear of Resurrection from the Dead. DOI : <http://dx.doi.org/10.13140/RG.2.2.34655.57768>

Tkemaladze, J. (2025). The Centriolar Theory of Differentiation Explains the Biological Meaning of the Centriolar Theory of Organismal Aging. Longevity Horizon, 1(3). DOI : <https://doi.org/10.5281/zenodo.15057288>

Tkemaladze, J. (2025). The Centriolar Theory of Differentiation Explains the Biological Meaning of the.

Tkemaladze, J. (2025). The Concept of Data-Driven Automated Governance. Georgian Scientists, 6(4), 399–410. DOI : <https://doi.org/10.52340/gS.2024.06.04.38>

Tkemaladze, J. (2025). The Stage of Differentiation Into Mature Gametes During Gametogenesis in Vitro. Longevity Horizon, 1(3). DOI : <https://doi.org/10.5281/zenodo.16808827>

Tkemaladze, J. (2025). The Tkemaladze Method Maps Cell Lineage with Mutant Mitochondrial Transfer. Longevity Horizon, 1(4). DOI : <https://doi.org/10.5281/zenodo.17236869>

Tkemaladze, J. (2025). The Tkemaladze Method: A Modernized Caucasian Technology for the Production of Shelf-Stable Activated Wheat with Enhanced Nutritional Properties. Longevity Horizon, 1(3). DOI : <https://doi.org/10.5281/zenodo.16905079>

Tkemaladze, J. (2025). Theory of Lifespan Decline. Longevity Horizon, 1(3). DOI : <https://doi.org/10.5281/zenodo.17142909>

Tkemaladze, J. (2025). Through In Vitro Gametogenesis — Young Stem Cells. Longevity Horizon, 1(3). DOI : <https://doi.org/10.5281/zenodo.15873113>

Tkemaladze, J. (2025). Tkemaladze, J. (2025). The Centriole Paradox in Planarian Biology: Why Acentriolar Stem Cells Divide and Centriolar Somatic Cells Do Not. Preprints. DOI : <https://doi.org/10.20944/preprints202509.0382.v1>

Tkemaladze, J. (2025). Unlocking the Voynich Cipher via the New Algorithmic Coding Hypothesis. Longevity Horizon, 1(3). DOI : <https://doi.org/10.5281/zenodo.17054312>

Tkemaladze, J. (2025). Uznadze Set Revisited. Longevity Horizon, 1(4). DOI : <https://doi.org/10.5281/zenodo.17609772>

Tkemaladze, J. (2025). Why do planarian cells without centrioles divide and cells with centrioles do not divide?. Longevity Horizon, 1(3). DOI : <https://doi.org/10.5281/zenodo.17054142>

Tkemaladze, J. (2025). Гаметогенез In Vitro: современное состояние, технологии и перспективы применения. Research Gate. DOI : <http://dx.doi.org/10.13140/RG.2.2.28647.36000>

Tkemaladze, J. (2026). The Strength of Clay, The Weakness of Gods. Longevity Horizon, 2(1). DOI : <https://doi.org/10.65649/2neyxv38>

Tkemaladze, J. (2026). Visions of the Future. Longevity Horizon, 2(1). DOI : <https://doi.org/10.65649/8be27s21>

Tkemaladze, J. Systemic Resilience and Sustainable Nutritional Paradigms in Anthropogenic Ecosystems. DOI : <http://dx.doi.org/10.13140/RG.2.2.18943.32169/1>

Tkemaladze, J. V., & Chichinadze, K. N. (2005). Centriolar mechanisms of differentiation and replicative aging of higher animal cells. Biochemistry (Moscow), 70, 1288-1303.

Tkemaladze, J., & Apkhazava, D. (2019). Dasatinib and quercetin: short-term simultaneous administration improves physical capacity in human. J Biomedical Sci, 8(3), 3.

Tkemaladze, J., & Chichinadze, K. (2005). Potential role of centrioles in determining the morphogenetic status of animal somatic cells. Cell biology international, 29(5), 370-374.

Tkemaladze, J., & Chichinadze, K. (2010). Centriole, differentiation, and senescence. Rejuvenation research, 13(2-3), 339-342.

Tkemaladze, J., & Gakely, G. (2025). A Novel Biotechnological Approach for the Production of Shelf-Stable, Nutritionally Enhanced Activated Wheat: Protocol Development, Nutritional Profiling, and Bioactivity Assessment. DOI : <https://doi.org/10.20944/preprints202508.1997.v1>

Tkemaladze, J., & Gakely, G. (2025). Induction of de novo centriole biogenesis in planarian stem cells. Longevity Horizon, 1(4). DOI : <https://doi.org/10.5281/zenodo.17283229>

Tkemaladze, J., & Samanishvili, T. (2024). Mineral ice cream improves recovery of muscle functions after exercise. Georgian Scientists, 6(2), 36–50. DOI : <https://doi.org/10.52340/gs.2024.06.02.04>

Tkemaladze, J., Gakely, G., Gegelia, L., Papadopulo, I., Taktakidze, A., Metreveli, N., ... & Maglakelidze, U. (2025). Production of Functional Gametes from Somatic Cells of the Planarian Schmidtea Mediterranea Via in Vitro Gametogenesis. Longevity Horizon, 1(3). DOI : <https://doi.org/10.5281/zenodo.17131291>

Tkemaladze, J., Tavartkiladze, A., & Chichinadze, K. (2012). Programming and Implementation of Age-Related Changes. In Senescence. IntechOpen.

Tkemaladze, Jaba and Kipshidze, Mariam, Regeneration Potential of the Schmidtea Mediterranea CIW4 Planarian. Available at SSRN: <https://ssrn.com/abstract=4633202> or <http://dx.doi.org/10.2139/ssrn.4633202>

Vorobjev, I. A., & Chentsov, Y. S. (1982). Centrioles in the cell cycle. I. Epithelial cells. *Journal of Cell Biology*, 93(3), 938–949. <https://doi.org/10.1083/jcb.93.3.938>

Winey, M., & O'Toole, E. (2014). Centriole structure. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 369(1650), 20130457. <https://doi.org/10.1098/rstb.2013.0457>

Wolf, D. P., Mitalipov, P. A., & Mitalipov, S. M. (2019). Mitochondrial replacement therapy in reproductive medicine. *Trends in Molecular Medicine*, 25(3), 215–226. <https://doi.org/10.1016/j.molmed.2018.12.004>

Yamashiro, C., Sasaki, K., & Saitou, M. (2018). Generation of human oogonia from induced pluripotent stem cells in vitro. *Science*, 362(6412), 356–360. <https://doi.org/10.1126/science.aat1674>

Yamashiro, C., Sasaki, K., Yabuta, Y., Kojima, Y., Nakamura, T., Okamoto, I., ... & Saitou, M. (2018). Generation of human oogonia from induced pluripotent stem cells in vitro. *Science*, 362(6412), 356–360. <https://doi.org/10.1126/science.aat1674>

Yamashiro, C., Sasaki, K., Yabuta, Y., Kojima, Y., Nakamura, T., Okamoto, I., Yokobayashi, S., Murase, Y., Ishikura, Y., Shirane, K., Sasaki, H., Yamamoto, T., & Saitou, M. (2018). Generation of human oogonia from induced pluripotent stem cells in vitro. *Science*, 362(6412), 356–360. <https://doi.org/10.1126/science.aat1674>

Zhang, Y., Liu, Y., & Sun, Q. Y. (2022). Centrosome regulation in female fertility. *Reproduction & Fertility*, 3(1), R1–R11. <https://doi.org/10.1530/RAF-21-0039>

Zhou, F., Wang, R., Yuan, P., Ren, Y., Mao, Y., Li, R., ... & Qiao, J. (2019). Reconstituting the transcriptome and DNA methylome landscapes of human implantation. *Nature*, 572(7771), 660–664. <https://doi.org/10.1038/s41586-019-1500-0>

Прангишвили, А. И., Гаситашвили, З. А., Мацаберидзе, М. И., Чичинадзе, К. Н., Ткемаладзе, Д. В., & Азмайпарашвили, З. А. (2017). К топологии антитеррористических и антикриминальных технологии для образовательных программ. В научном издании представлены материалы Десятой международной научно-технической конференции «Управление развитием крупномасштабных систем (MLSD'2016)» по следующим направлениям:• Проблемы управления развитием крупномасштабных систем, включая ТНК, Госхолдинги и Гос-корпорации., 284.

Прангишвили, А. И., Гаситашвили, З. А., Мацаберидзе, М. И., Чхартисвили, Л. С., Чичинадзе, К. Н., & Ткемаладзе, Д. В. (2017). & Азмайпарашвили, З.А (2017). Системные составляющие здравоохранения и инноваций для организации европейской нано-биомедицинской экосистемной технологической платформы. Управление развитием крупномасштабных систем MLSD, 365-368.

Ткемаладзе, Д. (2025). Асимметрия в наследовании centrosom/центриолей и ее последствия. DOI : <http://dx.doi.org/10.13140/RG.2.2.34917.31206>

Ткемаладзе, Д. (2025). Гаметогенез in vitro (IVG)-Этап дифференцировки в зрелые гаметы. DOI : <http://dx.doi.org/10.13140/RG.2.2.20429.96482>

Ткемаладзе, Д. (2025). Дифференциация соматических клеток многоклеточных животных. DOI : <http://dx.doi.org/10.13140/RG.2.2.23348.97929/1>

Ткемаладзе, Д. (2025). Индукция примордиальных клеток, подобных зародышевым клеткам (PGCLCs) современные достижения, механизмы и перспективы применения. DOI : <http://dx.doi.org/10.13140/RG.2.2.27152.32004>

Ткемаладзе, Д. (2025). Репликативный Лимит Хейфлика. DOI : <http://dx.doi.org/10.13140/RG.2.2.25803.30249>

Ткемаладзе, Д. (2025). Элиминация Центриолей: Механизм Обнуления Энтропии в Клетке. DOI : <http://dx.doi.org/10.13140/RG.2.2.12890.66248/1>

Ткемаладзе, Д. В., & Чичинадзе, К. Н. (2005). Центриолярные механизмы дифференцировки и репликативного старения клеток высших животных. Биохимия, 70(11), 1566-1584.

Ткемаладзе, Д., Цомаиа, Г., & Жоржолиани, И. (2001). Создание искусственных самоадаптирующихся систем на основе Теории Прогноза. Искусственный интеллект. УДК 004.89. Искусственный интеллект. УДК 004.89.

Чичинадзе, К. Н., & Ткемаладзе, Д. В. (2008). Центросомная гипотеза клеточного старения и дифференциации. Успехи геронтологии, 21(3), 367-371.

Чичинадзе, К., Ткемаладзе, Д., & Лазарашвили, А. (2012). Новый класс рнк и центросомная гипотеза старения клеток. Успехи геронтологии, 25(1), 23-28